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SYNTHESIS OF N-PHOSPHORYLATED AMINOACIDS

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Dedicated to Professor Reinhard Schmutzler on the occasion of his 60th birthday

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N-Phosphorylated α -aminoacids have been synthesized by the reaction of organophosphoryl chlorides with O,N-bis(trimethylsilylated) derivatives of the corresponding aminoacids.

Key words: N-Phosphorylated aminoacids, synthesis, O-silylated intermediates, NMR.

INTRODUCTION

One of the potential synthetic routes to prepare N-phosphorylated peptides embraces the preparation of N-phosphorylated aminoacids. It may be accomplished using the Todd-Atherton reaction of dialkylphosphites with α -aminoacids (Figure 1). Some limitations of this method are worthy of mention. Thus, as to dialkylphosphites, satisfactory results have been attained for diisopropylphosphite and dibutylphosphite only, whereas under the conditions of the Todd-Atherton reaction, dimethylphosphite gives rise to a complex mixture of compounds. In addition, the racemization of the aminoacid or its N-phosphorylated derivative can be provoked by the use of excess free base (3–4 moles of triethylamine per 1 mole of aminoacid).

This paper reports the synthesis of N-phosphorylated α -aminoacids via their N,O-bis(trimethylsilylated) derivatives, following the method developed earlier^{5,6} for the synthesis of α -acylaminocarboxylic⁵ and phosphonic acids.⁶ Additionally, we focused our attention on the study of NMR-spectra of these phosphoaminoacids.

(RO)₂PHO +
$$H_2$$
NCHCOOH $\xrightarrow{\text{CCl}_4,\text{Et}_3\text{N}}$ (RO)₂P(O)NHCHCOOH + CHCl₃

RESULTS AND DISCUSSION

N,O-Bis(trimethylsilylated) α -aminoacids 1a-c, prepared according to the literature, 7 were reacted with dialkylphosphorochloridates 2a,d, phosphonic chloride 2e or diphenylphosphinic chloride 2f at ambient temperature in acetonitrile for 1-3 hours to give trimethylsilyl esters of N-phosphorylated aminoacids (3a-g); Scheme 1). All compounds 3 are sensitive to atmospheric moisture. Compound 3a has been isolated analytically pure, its structure being characterized by 1H and ^{31}P -NMR

1-4	R ¹	R ²	RЗ	
a	Н	EtO	Bt0	
b	Me	EtO	EtO	
c	i-Pr	EtO	Et0	
đ	Ме	i-PrO	i-PrO	
е	Н	Me	Et0	
f	Me	Ph	Ph	
g	i-Pr	Ph	Ph	

SCHEME 1 Synthesis of N-phosphorylated aminoacids and their intermediate silylic esters.

spectral data (see Experimental). Ester 3a can be purified by distillation in vacuo⁸; compound 3e was decomposed when distilled. Compounds 3b-d and 3f,g were not isolated from the reaction mixtures, but were hydrolyzed directly under mild conditions to give 4b-d,f,g. Similarly, the compounds 3a,e have been hydrolyzed, without their preliminary isolation, to provide 4a,e. It should be pointed out that only volatile side-products are formed in both steps; thus, the purification of 4a-g is quite easy. The yields of acids 4a-d,g are high (81-98%), those of 4e,f are moderate (41 and 46%). The structures of 4a-g have been determined by NMR ¹H and ³¹P-spectral data and microanalysis. Phosphorylamidophosphate 4d has been prepared by the literature procedure²; both specimens had identical characteristics.

The phosphorylation of N-silylated derivatives is also a useful method for the

$$\begin{array}{c}
\text{Me}_3\text{Si-NH} & \text{CO}_2\text{SiMe}_3 \\
\text{S-SiMe}_3 & + 2\mathbf{d} & \\
\mathbf{8} & & \\
\end{array}$$

SCHEME 2 Synthesis of phosphorylated dipeptide 7 and cystein 10.

synthesis of N-phosphorylated dipeptides (7) or α -aminoacids, containing side-chain functional groups (10) (Scheme 2).

However, one limitation should be mentioned. When 2f is used as the phosphorylating agent, the rates of N- and O-phosphorylation are comparable, which is reflected by noticeable amounts of N-phosphorylated dipeptides formed by the interaction of 1b,c with intermediate mixed anhydrides. The yields of dipeptides, characterized as anilides, are relatively high (see Experimental).

The ¹H-NMR spectra of compounds **4b-d** are characterized by the double magnetic nonequivalence^{9,10} of the protons of alkoxy-groups at phosphorus. Two factors influence the multiplicity of methylenic protons of one CH₂O group in **4b-c** and every methyl group of isopropoxy-radicals of **4d**: the asymmetric center on the α -carbon atom of the amino-acid residue, which causes diastereotopy of alkoxy-groups, and conformational differences of groups. Compound **4b** has identical constants ${}^3J_{\text{CH}_3\text{CH}_2}$ and ${}^3J_{\text{POCH}_2}$; that result in quintet for one CH₂O-group and typical AB-system for the other one (${}^2J = 10 \text{ Hz}$). Two clear doublets (δ 1.27 and 1.24 ppm) can be observed for methyl groups of one isopropoxy radical of **4c**, as to the other radical, the diminished chemical shift difference between doublet centers can hardly be registered, the resolution of NMR-spectrometer being taken into consideration.

The magnetic non-equivalence of alkoxy-protons^{9,10} in ¹H-NMR spectra of 4b,d (see Experimental) being connected with the conformational flexibility of the alkoxy-group at phosphorus, the synthesis of some derivatives of 4 seemed reasonable to study the influence of such a derivatisation of structure on their spectral characteristics. Since some of the N-phosphorylated aminoacids 4 decomposed rapidly under storage, the stable anilides 12 have been synthesized via their mixed anhydrides¹¹ (Scheme 3). As to compound 14, prepared under conditions similar to that used for the synthesis of 12, the unique signal $\delta = 5.3$ ppm in its ³¹P-NMR spectrum clearly shows the absence of essential racemization in the course of N-phosphorylation.

The "double magnetic non-equivalence" mentioned above for 4d disappears in ¹H-NMR spectrum of 12d, non-equivalence of CH₃-groups caused by the chiral carbon atom being diminished to 0.015 ppm as compared with 4d ($\Delta \delta = 0.032$ ppm) at the same time. The reduced association of 12d in solution as well as the enhanced conformational flexibility of its isopropoxy groups as compared with 4d may be responsible for this effect. On the contrary, the second amino-acid fragment insertion results in the magnetic non-equivalence of all four methyl groups in isopropoxy-radicals of 14. It may be associated with the more rigid conformation of 14 as compared to 12d in view of intra- or intermolecular associative bonds. It is confirmed by the increased (0.119 ppm) chemical shift difference for methine protons of the isopropoxy groups of 14, this value being equal to 0.046 ppm for 4d and negligible for 12d. The CH3-decoupled signal of one methine proton turns out to be a doublet with ${}^{3}J_{POCH} = 7.6$ Hz, but the components of the second methine proton doublet remain split with ${}^5J_{NHPOCH} = 1.5$ Hz. The long distance coupling occurrence may be explained by the planar structure of C-O-P-Nfragment reported elsewhere¹² for the crystal structure of 4d. Such a spatial orientation of one isopropoxy group may limit the conformational flexibility of the molecule.

4a,d,g

11a,d,g

$$R^2$$
 R^3
 NH
 $CONHPh$

12a,d,g

N-Methylmorpholine/THF/DMF -15°C,1h--r.t.,3h

SCHEME 3 Synthesis of N-phosphorylated aminoacids anilides.

CONCLUSION

The reaction of O,N-bis(trimethylsilylated) derivatives of aminoacids with organophosphorylchlorides could be used for preparation of N-phosphorylated aminoacids and their derivatives, the structure of starting materials being taken into consideration.

EXPERIMENTAL

Methods. ¹H-NMR and ³¹P-NMR-spectra were recorded on a Bruker WP-200 spectrometer (200/81 MHz); chemical shifts of ¹H-NMR are reported in parts per million relative to internal hexamethyldisiloxane (0.00 ppm). 1% H₃PO₄ was used as external standard for ³¹P-NMR spectra. IR spectra were measured on a UR 20 spectrometer. Observed rotations at the Na-D line were obtained using a Polamat A polarimeter. The reactions were monitored and the purity of compounds was checked by thin-layer chromatography, using analytical TLC plates (Silufol UV₂s₄), purchased by Sklo Union (CSFR). Melting points were determined on a Boetius micro-mp apparatus and are not corrected. Elemental analyses were performed by the Department of Analytical Chemistry.

Preparation. All reagents were distilled freshly. Reagent grade solvents were purified before use. Acetonitrile was distilled over P₂O₅.

The preparation of silylated amino-acids **1a-c** and **8** was carried out according to the literature, ^{7,13} their structure being confirmed by ¹H-NMR spectroscopy. Starting materials **2** were synthesized as previously ¹⁴⁻¹⁷ described. All physical constants and spectroscopic data of compounds agree with the literature values.

Preparation of N,O-Bis(trimethylsilyl)glycyl-glycine, 5. Modified Procedure. To a vigorously stirred suspension of glycyl-glycine (0.76 g, 5.7 mmol) in acetonitrile (4 mL) was added trimethylsilyldiethylamine (2.17 g, 14.9 mmol). The mixture was stirred for 2 hrs until the dissolution is completed. The solvent was removed in vacuo to give 5 (1.61 g, 100%) as a colorless residue. H-NMR (CDCl₃, HMDS): 0.07 (s, 9H, Me₃SiN); 0.29 (s, 9H, Me₃SiO); 0.91 (t, 1H, J = 9.0 Hz, NHSi); 3.36 (d, 2H, J = 9.0 Hz, CH₂CON); 4.02 (d, 2H, J = 5.4 Hz, CH₂COO); 7.50 (t, 1H, J = 5.4 Hz, CONH). The so obtained 5 was used directly for further reaction with 2d.

N-(Diethoxyphosphoryl)-O-trimethylsilylglycine, **3a**. To the solution of **1a** (0.696 g, 3.17 mmol) in acetonitrile (1.5 mL) was added dropwise phosphorochloridate **2a** (0.546 g, 3.17 mmol) in acetonitrile (2 mL). The mixture was stirred at 20°C for 0.5 h, then allowed to stand for 2.5 hrs and evaporated at 40°C under reduced pressure to afford **3a** as colorless thick syrup. Pure **3a** (0.720 g, 80%) may be obtained by direct distillation of the crude product under protection from traces of moisture, b.p. 140–143°C/0.1 Torr. ¹H-NMR (CDCl₃/HMDS): 0.20 (s, 9H, Me₃Si), 1.21 (t, 6H, J = 7.0 Hz, CH₂CH₃), 3.36 (dt, 1H, J = 10.6, 6.2 Hz, NH), 3.57 (dd, 2H, $J_{PNCH_2} = 9.8$ Hz, CH₂CO), 3.98 (pseudo q, 4H, $J_{POCH_3} = 7.2$ Hz, CH₂CH₃). ³P-NMR (CDCl₃): $\delta = 7.9$ ppm. Anal. Calcd. for C₉H₂₂NO₃PSi (mol. weight 283.4), C, 38.15; H, 7.83; P, 10.93; Si, 9.91. Found: C, 38.12; H, 7.79; P, 11.10; Si, 10.34.

N-(O-Ethylmethylphosphonyl)-O-trimethylsilylglycine, **3e**. The workup was carried out by the procedure, described for ester **3a**. The residual colorless oil was allowed to stand for 1h at $40^{\circ}\text{C}/0.1$ Torr to afford crude **3e** of more than 97% purity, suitable for further reaction. ¹H-NMR (CDCl₃/HMDS): 0.28 (s, 9H, Me₃Si), 1.27 (t, 3H, J = 7.0 Hz, CH₂CH₃), 1.47 (d, 3H, J = 17.0 Hz, CH₃P), 3.64–3.83 (m, 2H, CH₂N), 3.96–4.10 (m, 3H, CH₂CH₃, NH). ³¹P NMR (CH₃CN): $\delta = 33.6$ ppm. Anal. Calcd. for C_xH₃₀NO₄PSi (mol. weight 252.3), C, 37.93; H, 7.96; P, 12.23; Si, 11.09. Found: C, 37.42; H, 7.82; P, 12.14; Si, 9.97.

The attempt to purify crude 3e by distillation (123-125°C/0.1 Torr) failed because of its rapid decomposition (see also 4e). Due to high lability of 3e its spectra include signals of corresponding acid 4e.

N-(Diethoxyphosphoryl)glycine, 4a. Typical procedure for water-soluble N-phosphorylated aminoacids. The reaction of 1a (1.59 g, 7.3 mmol) and 2a (1.14 g, 6.6 mmol) was carried out by the procedure described for 3a. After removal of the solvent with a rotary evaporator, crude 3a was chromatographed on a short silica gel column, eluted sequentially with neat CHCl₃ and CHCl₃/EtOH mixtures, to give 4a (1.37 g, 98%) as the thick colorless oil, which readily crystallized on cooling in the refrigerator, mp 36–38°C. 'H NMR (CDCl₃/HMDS): 1.30 (t, 6H, J = 7.1 Hz, 2CH₃), 3.64 (d, 2H, J = 4.7 Hz, NHC<u>H₂</u>), 4.05 (m, 4H, J_1 = 7.1 Hz, J_2 = 8.0 Hz, CH₂O), 5.42 (br s, 1H, NH), 9.98 (br s, 1H, COOH). ³¹P NMR (CDCl₃): δ = 7.7 ppm. Anal. Calcd. for C₆H₁₄NO₅P (mol. weight 211.1), C, 34.13; H, 6.68; N, 6.63; P, 14.67. Found: C, 34.41; H, 6.59; N, 6.50; P, 14.62.

N-(O-Ethylmethylphosphonyl)glycline, 4e. The reaction of 1a (0.46 g, 2.1 mmol) in acetonitrile (2.5 mL) and 2e (0.30 g, 2.1 mmol) in acetonitrile (1 mL) was carried out by the procedure described for 3e. After removal of the solvent under reduced pressure crude 3e was flash-chromatographed immediately on a short silica gel column, eluted sequentially with neat CHCl₃ and CHCl₃/EtOH mixtures to afford 4e (0.16 g, 41%) as a white solid. Recrystallisation from ethyl ether/ethanol gave pure 4e, mp $100-101^{\circ}$ C. ¹H NMR (CDCl₃/HMDS): 1.24 (t, 3H, J = 7.1 Hz, CH₃CH₂), 1.42 (d, 3H, J = 16.8 Hz, CH₃P), 3.55 (unresolved dd, 2H, CH₂), 3.82-4.07 (m, 2H, CH₂CH₂), 5.11 (br s, 1H, NH), 11.04 (br s, 1H, COOH). ³¹P NMR (CDCl₃): $\delta = 35.5$ ppm. Anal. Calcd. for C₅H₁₂NO₄P (mol. weight 181.1), C, 33.15; H, 6.67; P, 17.10. Found: C, 33.27; H, 6.78; P, 17.34. Prolonged storage of non-purified acid 4e is accompanied by its decomposition.

N-(Diethoxyphosphoryl)aminoacids, 4b,c. General Procedure. To a solution of 1b,c (3 mmol) in acetonitrile (3 mL) compound 2a (3 mmol) was added and the mixture was maintained at room temperature for 1-3 hrs. After removal of the solvent by evaporation in vacuo the residue was dissolved in 5 mL of dry ether and 3-4 drops of water were added. The mixture was evaporated in vacuo, and then the residue was dissolved in 20 mL CHCl₃ and washed with 10 mL of 10% aqueous solution of citric acid. The aqueous layer was extracted with chloroform (2 × 20 mL), the combined organic layers were dried over anhydrous Na₂SO₄. The solvent was evaporated under reduced pressure to afford 4b (83%) as a crystalline product, mp 74-76°C (CHCl₃-hexane), and 4c (86%) as a viscous oil, their purity being confirmed by TLC (EtOH/BuOH/NH₄OH/H₂O, 4:4:4:1).

N-(Diethoxyphosphoryl)-L-alanine, **4b**. ¹H NMR (CDCl₃/HMDS): 1.29 (t, 3H, J = 7 Hz, CH₃CH₂), 1.32 (t, 3H, J = 7 Hz, CH₃CH₂), 1.43 (d, 3H, J = 7 Hz, CH₃CH₂), 3.83 (m, 1H, $J_1 = 7$ Hz, $J_2 = 7.7$ Hz, $J_3 = 7.2$ Hz, NHCH), 4.03 (m, 1H, $J_1 = 7$ Hz, $J_2 = 7.1$ Hz, $J_3 = 10.1$ Hz, CHO), 4.10 (m, 1H, $J_1 = 7$ Hz, $J_2 = 7.1$ Hz, $J_3 = 10.1$ Hz, CHO), 4.04 (dq, 2H, $J_1 = 7$ Hz, $J_2 = 7.1$ Hz, CH₃CH₂O), 4.98 (dd, 1H, $J_1 = 7.7$ Hz, $J_2 = 12.5$ Hz, NH), 9.57 (br s, 1H, COOH). ³¹P NMR (CDCl₃): δ = 7.9 ppm. Anal. Calcd. for C₇H₁₆NO₅P × 0.5 H₂O (mol. weight 225.2), C, 35.90; H, 7.32; N, 5.98; P, 13.23. Found: C, 36.09, H, 7.17; N, 6.13; P, 13.46.

N-(Diethoxyphosphoryl)-D-valine, 4c. ¹H NMR (CDCl₃/HMDS): 0.81 (d, 3H, J = 6.8 Hz, CH₃CH), 0.89 (d, 3H, J = 6.8 Hz, CH₃CH), 1.18 (t, 3H, J = 7 Hz, CH₃CH₂), 1.20 (t, 3H, J = 7 Hz, CH₃CH₂), 2.00 (m, 1H, $J_1 = 6.8$ Hz, $J_2 = 4.6$ Hz, CH₃CH₂CH₃), 3.51 (m, 1H, $J_1 = 4.6$ Hz, $J_2 = 10.4$ Hz, $J_3 = 10.4$ Hz, NHCH), 3.81 (dd, 1H, $J_1 = 10.4$ Hz, $J_2 = 10.4$ Hz, NH), 3.95 (m, 1H, $J_1 = 7$ Hz, $J_2 = 7$ Hz, $J_3 = 10.2$ Hz, CHO), 3.98 (m, 1H, $J_1 = 7$ Hz, $J_2 = 7$ Hz, $J_3 = 10.2$ Hz, CHO), 3.96 (m, 2H, $J_1 = 7$ Hz, $J_2 = 7$ Hz, $J_3 = 7$ Hz, CH₂O), 11.47 (br s, 1H, COOH). ³¹P NMR (CDCl₃): δ = 8.5 ppm. Anal. Calcd. for C₉H₂₀NO₅P (mol. weight 253.2), C, 42.69; H, 7.96; P, 12.23. Found: C, 42.20; H, 7.88; P, 11.85.

N-(Diisopropoxyphosphoryl)-L-alanine, **4d**. *Method a*: To a solution of **2d** (1.04 g, 5.2 mmol) in actonitrile (6 mL) was added **1b** (1.61 g, 6.9 mmol), and the mixture was maintained at room temperature for 1 h. After removal of the solvent under reduced pressure, the residue was treated as described for **4b**,c to afford crude **4d**. Recrystallisation from chloroform/hexane gave pure **4d** (1.16 g, 88%), mp 118–119°C, [α]_D¹² – -6° ± 0.2° (c = 5.0, C₂H₅OH) (lit.² mp 110–2°C, [α]_D¹⁴ – -7° (c = 1, C₂H₅OH)). ¹H NMR (CDCl₃/HMDS): 1.24 (d, 3H, J = 6.5 Hz, CH₃CHCH₃), 1.27 (d, 3H, J = 6.4 Hz, CH₃CHCH₃), 1.29 (d, 6H, J = 6.2 Hz, CH₃CHCH₃), 1.40 (d, 3H, J = 7 Hz, CH₃CHNH), 3.80 (m, 1H, J₁ = 7 Hz, J₂ = 8 Hz, J₃ = 7.5 Hz, NHCH₃), 4.54 (m, 1H, J₁ = 6.5 Hz, J₂ = 7.5 Hz, CHO), 4.58 (m, 1H, J₁ = 6.2 Hz, J₂ = 7.5 Hz, CHO), 4.73 (dd, 1H, J₁ = 8 Hz, J₂ = 12.2 Hz, NH₂), 11.31 (br s, 1H, COOH). ³¹P NMR (CDCl₃): δ = 6.1 ppm. Anal. Calcd. for C₉H₂₀NO₅P (mol. weight 253.2), C, 42.69; H, 7.96; N, 5.53; P, 12.23. Found: C, 42.58; H, 8.05; N, 5.50; P, 12.38.

Method b: The reaction of diisopropylphosphite (1.66 g, 10 mmol) and **1b** (0.89 g, 10 mmol) was accomplished as described earlier² to afford crude **4d** (2.29 g, 90%). The crude **4d** was purified by crystallisation (CHCl₃-hexane) to give authentic **4d**, mp 104–107°C.

N-(Diphenylphosphinyl)-D-alanine, **4f**. To solution of **1b** (D-isomer, 1.21 g, 5.2 mmol) in acetonitrile (3.0 mL) was added a solution of **2f** (1.12 g, 4.7 mmol) in acetonitrile (2.0 mL). The solution was immediately evaporated, the residual light-yellow oil was dissolved in 40 mL of EtOAc, washed with 20 mL of water, 10% aqueous solution of citric acid (2 \times 20 mL), 20 ml of water and 20 mL of brine.

The organic layer was dried over anhydrous Na₂SO₄, filtered and evaporated under reduced pressure. The residual white glass was purified by chromatography on a silica gel column, eluted with mixture of hexane/CHCl₃/EtOH (50:50:4), to give white solid. Recrystallisation from chloroform/hexane gave pure 4f (0.57 g, 46%), mp 141–143°C, $[\alpha]_D^{13} + 20.3 \pm 1.5^{\circ}$ (c = 1.03, MeOH) (lit. ¹⁸ for L-isomer of 4f $[\alpha]_D^{25} - 21.4^{\circ}$ (c = 1.0, MeOH). ¹H NMR (CDCl₃/HMDS): 1.37 (d, 3H, J = 6.8 Hz, CH₃), 3.96 (dd. 1H, $J_1 = 5.6$ Hz, $J_2 = 10.4$ Hz, NH), 3.81 (m, 1H, $J_1 = 6.8$ Hz, $J_2 = 10.4$ Hz, $J_3 = 10.3$ Hz, NHCH), 7.35–7.96 (m, 10 H_{arom}), 12.61 (br s, 1H, COOH). ³¹P NMR (CDCl₃): $\delta = 26.5$ ppm. Anal. Calcd. for C₃H₁₆NO₃P (mol. weight 289.3), C, 62.28; H, 5.57; N, 4.84; P, 10.71. Found: C, 62.01; H, 5.64; N, 4.61; P, 10.58.

N-(*Diphenylphosphinyl*)-*D-valine*, **4g**. The reaction of **1c** (1.69 g, 6.5 mmol) and **2f** (1.44 g, 6.1 mmol) was carried out by the procedure described for **4f**, to afford **4g** (1.54 g, 81%), as a white glass, $[\alpha]_{0}^{25}$ + 22.8 ± 1.4° (c = 1.14, MeOH) (lit. ¹⁸ for L-isomer of **4g** $[\alpha]_{0}^{25}$ - 15.2 (c = 1.0, MeOH)). ¹H NMR (CDCl₃/HMDS): 0.89 (d, 3H, J = 6.8 Hz, CH₃), 0.92 (d, 3H, J = 6.8 Hz, CH₃), 2.06 (m, 1H, J = 6.8 Hz, CH), 3.47–3.81 (m, 2H, NHCH), 7.43–7.95 (m, 10H_{urom}), 11.16 (br s, 1H, COOH). ³¹P NMR (CDCl₃): δ = 27.1 ppm. Anal. Calcd. for C₁₇H₂₀NO₃P (mol. weight 317.3), C, 64.35; H, 6.35; N, 4.41; P, 9.76. Found: C, 64.39; H, 6.38; N, 4.18; P, 9.39.

N-(Diisopropoxyphosphoryl)glycyl-glycine, 7 and N-(diisopropoxyphosphoryl)-L-cysteine, 10. The reaction of 5 (1.61 g, 5.7 mmol) and 2d (1.08 g, 5.4 mmol), or 8 (1.37 g, 4.1 mmol) and 2d (0.77 g, 3.8 mmol), was carried out by the procedure described for 4a, to give 7 (97%) and 10 (87%).

Compound 7: mp 111–113°C (CHCl₃/hexane). ¹H NMR (CDCl₃/HMDS): 1.29 (d, 6H, J = 5.9 Hz, 2CH₃), 1.31 (d, 6H, J = 5.9 Hz, 2 CH₃), 3.62 (d, 2H, J = 11.1 Hz, CH₂NP), 4.12 (d, 2H, J = 4.9 Hz, CH₂NCO), 4.35 (br s, 1H, NHP), 4.58 (m, 2H, $J_1 = 5.9$ Hz, $J_2 = 8.2$ Hz, CHO), 7.30 (br t, 1H, J = 4.9 Hz, CONHCH₂), 10.43 (br s, 1H, COOH). ³¹P NMR (CDCl₃): $\delta = 6.2$ ppm. Anal. Calcd. for C₁₀H₂₁N₂O₆P (mol. weight 296.3), C, 40.54; H, 7.14; N, 9.46; P, 10.46. Found: C, 40.40; H, 7.34; N, 9.30; P, 10.43.

Metastable form of 7, mp 81-83°C, was also isolated (lit19 mp 73.5-74.5°C).

Compound 10: oil (lit.³⁰ oil). ¹H NMR (CDCl₃/HMDS): 1.28 (d, 3H, J = 6.2 Hz, CH₃), 1.31 (d, 3H, J = 6.2 Hz, CH₃), 1.32 (d, 3H, J = 6.2 Hz, CH₃), 1.33 (d, 3H, J = 6.2 Hz, CH₃), 1.63 (m, 1H, $J_1 = 7.7$ Hz, $J_2 = 9.7$ Hz, SH), 2.89 (m, 1H, $J_1 = 7.7$ Hz, $J_2 = 5$ Hz, $J_3 = 13.6$ Hz, CHS), 2.95 (m, 1H, $J_1 = 9.7$ Hz, $J_2 = 3.5$ Hz, $J_3 = 13.6$ Hz, CHS), 4.09 (m, 1H, $J_1 = 5$ Hz, $J_2 = 3.5$ Hz, $J_3 = 8.6$ Hz, $J_4 = 8.2$ Hz, NHCH), 4.54 (dd, 1H, $J_1 = 8.6$ Hz, $J_2 = 11.8$ Hz, NH), 4.60 (m, 1H, $J_1 = 6.2$ Hz, $J_2 = 7$ Hz, CHO), 4.64 (m, 1H, $J_1 = 6.2$ Hz, $J_2 = 7$ Hz, CHO), 10.27 (br s, 1H, COOH). ³¹P NMR (CDCl₃): δ = 5.8 ppm.

The compound 7 was obtained also by the literature² method, yield 76%.

N-(Diisopropoxyphosphoryl)glycyl-glycine anilide. To a solution of 7 (1.20 g, 4.05 mmol) and aniline (0.38 g, 4.05 mmol) in dry methylene chloride (20 mL) was added a 1.02 M solution of DCC in methylene chloride (4.05 mL, 4.13 mmol) at -20° C. The mixture was placed immediately in the refrigerator (0°C) and maintained at this temperature for 2 hrs. Then the reaction mixture was stirred at room temperature for 4 hrs. Upon addition of 5 drops of acetic acid, the precipitated dicyclohexylurea was removed by filtration and the solution was evaporated in vacuo. The residue was dissolved in EtOAc (20 mL), washed with 10 mL of 10% aqueous solution of citric acid, 10 mL of water, 10 mL of 10% aqueous NaHCO₃ and 10 mL of brine. The organic layer was dried over anhydrous Na₂SO₄, filtered and evaporated in vacuo to afford crude anilide. Recrystallisation from chloroform/hexane gave N-(diisopropoxyphosphoryl)glycyl-glycine anilide (1.08 g, 72%), as colorless solid, mp 118–120°C. 'H-NMR (CDCl₃/HMDS): 1.24 (d, 6H, J = 6.1 Hz, CH₃CHCH₃), 3.63 (d, 2H, J = 12.4 Hz, CH₂NP), 4.10 (d, 2H, J = 5.5 Hz, CH₂NC), 4.24 (br s, 1H, NHP), 4.56 (m, 2H, $J_1 = 6.1$ Hz, $J_2 = 7.4$ Hz, CHO), 7.03 (t, 1H_{arom}, J = 7.3 Hz), 7.23 (dd, 2H_{arom}, J = 7.3 Hz, $J_2 = 7.9$ Hz), 7.53 (d, 2H_{arom}, J = 7.9 Hz), 7.90 (t, 1H, J = 5.5 Hz, CONHCH₂), 9.29 (s, 1H, NHPh). 'P-NMR (CDCl₃): $\delta = 6.5$ ppm. Anal. Calcd. for C₁₆H₂₆N₃O₅P (mol. weight 371.4), C, 51.75; H, 7.06; N, 11.31; P, 8.34. Found: C, 51.49; H, 7.02; N, 11.23; P, 8.21.

N-(Diisopropoxyphosphoryl)-L-alanine anilide, 12d. A solution of isobutylchloroformate (0.27 g, 2 mmol) in tetrahydrofurane (6 mL) was added dropwise to a solution of 4d (0.51 g, 2 mmol) and N-methylmorpholine (0.20 g, 2 mmol) in tetrahydrofurane (11 mL) at -15° C under stirring. The mixture was stirred for 10 min at the same temperature, then the solution of aniline (0.19 g, 2 mmol) in tetrahydrofurane (3 mL) was added. Stirring was continued for 1 h at -15° C and 2 hrs at room temperature. An equal volume of water was added, the resulting mixture was concentrated, the precipitate was collected by filtration, then dissolved in CHCl₃, washed with aqueous citric acid, saturated aqueous NaHCO₃, water and dried over anhydrous Na₂SO₄. Removal of the solvent afforded solid crude 12d; recrystallisation from chloroform/hexane gave pure 12d (0.5 g, 77%), mp 151-152.5°C,

[α] $_{0}^{20.5}$ - 61.4 \pm 0,7° (c = 1.0, CHCl₃). ¹H NMR (CDCl₃/HMDS): 1.28 (d, 9H, J = 6 Hz, CH₃CHCH₃), 1.30 (d, 3H, J = 6 Hz, CH₂CHCH₃), 1.47 (d, 3H, J = 6.5 Hz, CH₃CHNH), 3.84 (m, $\overline{1}$ H, J_1 = 9.7 Hz, $\overline{2}^{11}$ J_2 = 10 Hz, $\overline{2}^{11}$ NHCH), 4.02 (m, J_1 = 6.5 Hz, J_2 = 10 Hz, $\overline{2}^{11}$ \overline{J}_3 = 10.8 Hz, $\overline{2}^{11}$ NHCH), 4.60 (m, 2H, J_1 = 6 Hz, J_2 = 7.5 Hz, $\overline{2}^{11}$ CHO), 7.06 (t, $\overline{1}$ H_{arom}, J = 7.4 Hz), 7.28 (dd, $\overline{2}$ H_{arom}, J_1 = 7.4 Hz, J_2 = 7.8 Hz), 7.61 (d, $\overline{2}$ H_{arom}, J = 7.8 Hz), 9.27 (br s, 1H, NHPh). ³¹P NMR (CDCl₃): δ = 5.5 ppm. Anal. Calcd. for C₁₅H₂₆N₂O₄P (mol. weight 328.4), C 54.87; \overline{H} , 7.67; N, 8.53; P, 9.43. Found: C, 54.63; H, 7.46; N, 8.44; P, 9.39.

N-(*Diethoxyphosphoryl*)glycine anilide, **12a**. The reaction of **4a** (0.94 g, 4.46 mmol) and isobutyl-chloroformate (0.58 g, 4.24 mmol) in the presence of N-methylmorpholine (0.45 g, 4.46 mmol), and with aniline (0.39 g, 4.24 mmol) was carried out in the same manner as for **12d**. Recrystallisation from chloroform/hexane gave **12a** (1.11 g, 92%), mp 93–94°C. 'H NMR (CDCl₃/HMDS): 1.31 (t, 6H, J = 7.1 Hz, CH₃), 3.78 (d, 2H, J = 12.1 Hz, NHCH₂), 4.00 (br s, 1H, NHP), 4.09 (m, 4H, $J_1 = 7.1$ Hz, $J_2 = 7.8$ Hz, CH₂O), 7.09 (t, 1H_{arom}, J = 7.4 Hz), 7.30 (dd, 2H_{arom}, $J_1 = 7.4$ Hz), 7.57 (d, 2H_{arom}, J = 8.1 Hz), 9.05 (br s, 1H, NHPh). ³¹P NMR (CDCl₃): $\delta = 8.3$ ppm. Anal. Calcd. for C₁₂H₁₉N₂O₄P (mol. weight 286.3), C, 50.35; H, 6.69; N, 9.79; P, 10.82. Found: C, 50.13; H, 6.61; N, 10.02; P, 10.62.

N-(Diphenylphosphinyl)-D-valine anilide, 12g, was obtained, following the same procedure; recrystal-lisation from chloroform/hexane gave 12g (75%), mp 194–197°C, [α] $_D^{21}$ +157.9° ± 0.3° (c = 1.0, CHCl₃). ¹H NMR (CDCl₃/HMDS): 0.95 (d, 3H, J = 6.5 Hz, CH₃), 0.98 (d, 3H, J = 6.3 Hz, CH₃), 2.40 (m, 1H, CH₂CHCH₃), 3.56–3.85 (m, 2H, NHCH), 7.04 (t, 1H_{arom}, J = 7.3 Hz), 7.25 (dd, 2H_{arom}, J = 7.3 Hz, J₂ = 7.9 Hz), 7.58 (d, 2H_{arom}, J = 7.9 Hz), 7.37–7.51 (m, 10H, Ph₂P), 9.96 (br s, 1H, NHPh). ³¹P NMR (CDCl₃): δ = 25.9 ppm. Anal. Calcd. for C₂₃H₂₅N₂O₂P (mol. weight 392.4), C, 70.39; H, 6.42; N, 7.14; P, 7.89. Found: C, 70.50; H, 6.48; N, 6.95; P, 7.83.

N-(Diphenylphosphinyl)-L-Valine anilide and N-(diphenylphosphinyl)-L-valyl-L-valine anilide. The reaction of O,N-bis(trimethylsilyl)-L-valine (1.66 g, 6.3 mmol) and 2f (1.42 g, 6.0 mmol) and the following workup were carried out according to procedure, described for 4f; 1.82 g of N-diphenylphosphinyl-L-valine (mixture with N-(diphenylphosphinyl)-L-valyl-L-valine) as white glass was obtained. This product was used on the second step without additional workup. To the stirred solution of glass obtained (0.62 g) and N-methyl-morpholine (0.20 g, 2.0 mmol) in THF (15 mL) a solution of isobutylchloroformate (0.27 g, 2.0 mmol) in THF (3.0 mL) was added at $-10 \div -15^{\circ}$ C. After 10 min the solution of aniline (0.18 g, 2.0 mmol) in THF (2 mL) was added and the mixture was stirred at - 10°C for 1 h and at +20°C for 2 hrs. Then the reaction mixture was evaporated to dryness and the residue was distributed between water (20 mL) and EtOAc (40 mL). The organic layer was washed by water (2 \times 15 mL), 10% aqueous citric acid (2 \times 15 mL), 1 M solution of NaHCO₃ (2 \times 15 mL) and brine (2 × 15 mL), dried over anhydrous Na₂SO₄ and evaporated. The residue (0.75 g) was crystallized fractionally from chloroform/hexane to give 0.327 g of N-(diphenylphosphinyl)-L-valine anilide, mp $194-197^{\circ}$ C, $[\alpha]_{1}^{17}-163.0^{\circ}\pm0.4^{\circ}$ (c = 1.0, CHCl₃), and 0.045 g of N-(diphenylphosphinyl)-L-valyl-Lvaline anilide, mp 284–286°C (dec). N-(Diphenylphosphinyl)-L-valine anilide. ¹H-NMR (CDCl₃/HMDS): 0.95 (d, 3H, J = 7.1 Hz, CH₃), 0.99 (d, 3H, J = 7.2 Hz, CH₃), 2.43 (m, 1H, CH₃CHCH₃), 3.53-3.75 (m, 2H, NHCH), 7.05-7.89 (m, $15H_{arom}$), 9.89 (br s, 1H, PhNH). ³¹P-NMR (CDCl₃): $\delta = 26.7$ ppm. Anal. Calcd. for C₂₃H₂₅N₂O₂P (mol. weight 392.4), C, 70.39; H, 6.42; N, 7.14; P, 7.89. Found: C, 70.35; H, 6.52; N, 6.99; P, 7.79. N-(Diphenylphosphinyl)-L-valyl-L-valine anilide. H-NMR (CDCl₃/ HMDS): = 0.93 (d, 3H, J = 6.9 Hz, CH_3 CHCH₃), 1.02 (d, 6H, J = 6.9 Hz, CH_3 CHCH₃), 1.09 (d, 3H, J = 6.9 Hz, CH_3CHCH_3 , 2.5 (m, $2\overline{H}$, $C\underline{H}(CH_3)_2$) 3.25 (dd, 1H, $J_1 = 9.6$ Hz, $\overline{J_2} = 9.7$ Hz, NHP), 3.53 (m, 1H, $J_1 = 9.6$ Hz, $J_2 = 6.8$ Hz, $J_3 = 3.5$ Hz, PNCH), 4.60 (dd, 1H, $J_1 = 9.8$ Hz, $J_2 = 4.3$ Hz, CONCH), 7.08-8.00 (m, 15 H_{arom}), 7.88 (d, 1H, J = 9.8 Hz, CONH), 9.86 (br s, 1H, PhNH). ³¹P-NMR (CDCl₃): $\delta = 25.99$ ppm. Anal. Calcd. for C₂₈H₃₄N₃O₃Px 0.25H₂O (mol. weight 491.6), C, 67.79; H, 7.0; P, 6.24. Found: C, 67.87; H, 7.09; P, 6.38.

L-Phenylalanine anilide hydrobromide, **13**. N-Carbobenzoxy-L-phenylalanine anilide³² was treated with 4 N HBr to give **13** as colorless solid; yield 95%; mp 228–231°C, $[\alpha]_{10}^{22}$ +95 ± 3° (c = 1.4, C₂H₅OH). Anal. Calcd. for C₁₅H₁₆N₂O' HBr (mol. weight 321.2), C, 56.09; H, 5.33; N, 8.72. Found: C, 55.89; H, 5.15; N, 8.66.

N-(Diisopropoxyphosphoryl)-L-alanyl-L-phenylalanine anilide, 14. A solution of isobutylchloroformate (0.27 g, 2 mmol) in THF (5 mL) was added dropwise to a solution of 4d (0.51 g, 2 mmol) and N-methylmorpholine (0.2 g, 2 mmol) in THF (15 mL) at -15° C with stirring. After stirring this mixture at -15° C for 10 min, a solution of 13 (0.64 g, 2 mmol) and N-methylmorpholine (0.2 g, 2 mmol) in DMF (7 mL) cooled to 0°C was added dropwise. The mixture was stirred at -15° C for 1 h and at room

temperature for 3 hrs. To obtain pure 14, the same procedure was applied as for the preparation of 12d. Recrystallisation from chloroform/hexane gave pure 14 (0.91 g, 97%), mp 230°C (dec.). 'H NMR $(CDCl_3/HMDS)$: 1.23 (d, 3H, J = 6.7 Hz, $C\underline{H}_3CHCH_3$), 1.24 (d, 3H, J = 6.4 Hz, $CH_3CHC\underline{H}_3$), 1.26 (d, 3H, J = 6.5 Hz, CH₃CHCH₃), 1.27 (d, 3H, J = 6.5 Hz, CH₃CHCH₃), 1.31 (d, 3H, J = 7.1 Hz, ²³ CH₃CHNH), 3.18 (m, 1H, $J_1 = 7.5$ Hz, $J_2 = 14$ Hz, CHPh), 3.24 (m, 1H, $J_1 = 6.1$ Hz, $J_2 = 14$ Hz, CHPh), 3.54 (dd, 1H, $J_1 = J_2 = 7.5$ Hz, NHP), 3.88 (m, 1H, $J_1 = 7.1$ Hz, 23 $J_2 = 7.5$ Hz, $J_3 = 9.7$ Hz.²³ CHNP), 4.46 (m, 1H, $J_1 = 6.5$ Hz, $J_2 = 7.5$ Hz, CHO), 4.58 (m, 1H, $J_1 = 6.5$ Hz, $J_2 = 7.5$ Hz, CHO), 4.94 (m, 1H, $J_1 = 6.1$ Hz, $J_2 = 7.5$ Hz, $J_3 = 9.1$ Hz, $J_3 = 9.1$ Hz, $J_4 = 6.1$ Hz, $J_5 = 7.5$ Hz, $J_5 = 7.5$ Hz, $J_7 = 7.5$ 7.4 Hz), 7.27 (dd, $2H_{arom}$, $J_1 = 7.4$ Hz, $J_2 = 7.7$ Hz), 7.59 ((d, $2H_{arom}$, J = 7.7 Hz), 7.25 (m, $5H_{arom}$ (Phe)), 7.63 (d, 1H, J = 9.1 Hz, CONHCH), 9.12 (s, 1H, NHPh). ³¹P NMR (CDCl₃): $\delta = 5.3$ ppm. Anal. Calcd. for C₂₄H₃₄N₃O₅P (mol. weight 475.5), C, 60.62; H, 7.21; N, 8.84; P, 6.51. Found: C, 60.83; H, 7.38; N, 8.68; P, 6.40. IR (KBr): 3288 (N—H), 1695, 1654, 1628, 1610, 1563 (CO—N—H), 1230 (P=O), 1029, 998 (P-O) cm⁻¹.

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REFERENCES AND NOTES

- 1. A. Cosmatos, I. Photaki and L. Zervas, Chem. Ber., 94, 2644 (1961).
- 2. G.-I. Ji, C.-B. Xue, J.-N. Zeng, L.-P. Li, W.-G. Chai and Y.-F. Zhao, Synthesis, 444 (1988).
- 3. X.-B. Ma and Y.-F. Zhao, J. Org. Chem., 54, 4005 (1989).
- 4. D. S. Kemp, in: "The Peptides. Analysis, Synthesis, Biology," (Academic Press, New York, 1979) eds. E. Gross and J. Meienhofer, Vol. I, p. 317.
- 5. D. R. Bolin, I.-I. Sytwu, F. Humiec and J. Meienhofer, Int. J. Peptide Protein Res., 33, 353 (1989).
- 6. V. Solodenko, T. Kasheva and V. Kuchar, Synth. Commun., 21, 1631 (1991).
- 7. K. Ruhlmann, Chem. Ber., 94, 1876 (1961).
- 8. A. B. Ouryupin, V. Yu. Komissarov, P. V. Petrovskii, Yu. A. Davidovich, T. A. Mastryukova and M. I. Kabachnik, Izv. RAN, Ser. Khim., 1694 (1992).
- 9. M. L. Martin, R. Mantione and G. L. Martin, *Tetrahedron Lett.*, 6, 3873 (1966). 10. E. I. Goryunov, P. V. Petrovskii, Y. Yu. Kudryavtsev, L. S. Zaharov and M. I. Kabachnik, *Dokl.* Akad. Nauk., (USSR), 281, 1378 (1985)
- 11. G. W. Anderson, J. E. Zimmerman and F. M. Callahan, J. Am. Chem. Soc., 83, 5012 (1967).
- 12. C. B. Xue, Y.-W. Yin, Y.-M. Liu, N.-J. Zhu and Y.-F. Zhao, Phosphorus, Sulfur and Silicon, 42, 149 (1989).
- 13. S. V. Rogozhin, Yu. A. Davidovich, S. M. Andreev, N. V. Mironova and A. I. Yurtanov, Izv. Akad. Nauk SSSR, Ser. Khim., 1868 (1974).
- 14. T. Mukayama and T. Fujisawa, Bull. Chem. Soc. Japan, 34, 812 (1961).
- 15. H. McCombie, B. S. Saunders and G. T. Stacey, J. Chem. Soc., 388 (1945).
- 16. B. Gallencamp, W. Hoper, B.-W. Kruger, F. Maurer and I. Pfister, in "Methoden der Organischen Chemie (Houben-Weil)" (Georg Thieme Verlag, Stuttgart-New York, 1982), ed. M. Regitz, Vol. E2, p. 337.
- 17. W. A. Higgins, P. W. Vogel and W. R. Craig, J. Am. Chem. Soc., 77, 1864 (1955).
- 18. R. Ramage, D. Hopton, M. J. Parrot, G. W. Kenner and G. A. Moore, J. Chem. Soc., Perkin Trans. I, 1357 (1984).
- 19. Y.-F. Zhao, D.-Q. Zhang and C. B. Xue, Int. J. Peptide Protein Res., 37, 457 (1991).
- 20. I.-C. Li and Y.-F. Zhao, *Phosphorus, Sulfur and Silicon*, **60**, 233 (1991).
- 21. Recorded at -54°C
- 22. D. W. Santi and P. V. Danenberg, *Biochemistry*, **10**, 4813 (1971).
- 23. CD₃OD was added.